Capnocytophaga canimorsus endocarditis with root abscess in a patient with a bicuspid aortic valve

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Introduction

Infective endocarditis caused by a zoonotic microorganism is a rare clinical condition. Capnocytophaga canimorsus is a bacterium of the normal oral flora of dogs and cats. It can be transmitted to humans by bite, scratch or others close contacts with dogs, and occasionally cats. It has been implicated in a spectrum of disorders including septicaemia, meningitis and septic arthritis. Splenectomy and alcoholism have been reported to be risk factors, but infection may occur in previously healthy patients.

We reported the case of a 65-year-old man with bicuspid aortic valve endocarditis and multiple abscesses of the aortic wall caused by the canine bacteria Capnocytophaga canimorsus. This case highlights the capacity of Capnocytophaga canimorsus to produce endocarditis whose incidence may be underestimated due to slow growth and incorrect identification of the organism.

Case report

A 65-year-old man was referred to our hospital for suspicion of endocarditis. His medical history included a cardiac follow-up for asymptomatic significant aortic stenosis. He adopted an antibiotic prophylaxis by amoxicillin (3g per os in the hour before a risk procedure). Cardiovascular risk factors were nicotinism, dyslipidemia and hypertension. He did not present other relevant pathologies. For two weeks, the patient presented intermittent fever associated with major asthenia. On initial examination, findings were fever at 39°C, tachycardia and 3/6 systolic murmur in the aortic area. Laboratory investigation revealed increased inflammatory indexes (C reactive protein: 175 ng/mL; white blood cells: 19x10⁹/L/mm³ with 85% neutrophils), normocytic anemia (10.5 g/dL), renal insufficiency (creatininemia: 150 micromoles/L, clearance: 43 mL/min). Transthoracic echocardiogram revealed a high suspicion of aortic endocarditis with vegetation. As the aortic valve was calcified and given the echogenicity, the analysis of the lesion was limited and not optimal. We then realized a transoesophageal echocardiogram for better delineation of the pathology and to exclude an extension to the other valves. Transoesophageal echocardiography showed a calcified bicuspid aortic valve with a vegetation on the posterior hemi valve (8 x 9 mm) and large abscesses affecting left coronary sinus, and the aortic root (Figure 1A and B). A multidetector computed tomographic angiography (MCTA) confirmed on 3D volume rendering showed three additional images on the aortic root, behind the main left coronary artery suggesting mycotic aneurysm or abscesses (Figure 1C). There was no significant lesion on the coronary arteries. Blood cultures were negative. The patient started an empirical parenterally antibiotic therapy with gentamycin (160 mg/24 h) plus ampicillin (12 g/24 h then 6 g/24 h after deterioration of creatinin clearance secondary to gentamycin and contrast perfusion during MCTA). He rapidly became afebrile and the inflammatory syndrome decreased. After 25 days, three anaerobia sets of blood cultures (chocolate agar plate incubated in a carbon dioxide-enriched environment) grew a Gram negative bacillus, oxidase and catalase positive, that was identified as Capnocytophaga species. The identification of Capnocytophaga canimorsus was made using 16S rDNA gene sequencing. The patient revealed he had four dogs at home. He did not report any bite but he made a habit of kissing these dogs. The patient underwent aortic valve and root replacement (intervention of Bentall) with the implantation of a valve conduit (Carbomedics n. 23). Intraoperatively, the surgical view confirmed the existence of three abscesses located in the left coronary sinus around the left coronary ostium and corroborated the ultra sound and MCTA findings. The patient had an uneventful post-operative recovery. He was discharged after a total of six weeks antibiotic treatment based on susceptibility test (organism sensitive for ampicillin).

Discussion

Capnocytophaga canimorsus is a commensal gram negative bacillus living in the saliva of dogs and cats. It is known to cause severe infection, mainly septicemia, meningitis, septic arthritis, and more rarely, infective endocarditis with less than 15 cases in the published literature since its identification in 1976.1 Capnocytophaga canimorsus is transmitted though bites from dogs (rarely cats) and close contacts with animals as in our observation. Significant risk factors for Capnocytophaga canimorsus infection are immunocompromised states including asplenism or chronic alcoholism.1 However, 40% of septicemia occurred in patients with no classical predisposing conditions.2 For infective endocarditis, underlying cardiac function...
condition at risk for endocarditis seems to be the most important risk factor. One third of cases of endocarditis had pre-existing cardiac lesions including mitral leaflet prolapse, aortic stenosis, mechanical valve, myxoma, heart murmur. Isolation of this slow-growing bacterium is difficult. Blood cultures (aerobic or anaerobic flasks) may be negative or the latter positive. The use of specific culture media (agar chocolate, brain heart infusion), CO2 and enriched atmosphere enhanced growth. PCR performed on the valve was recently used with success for the diagnosis of Capnocytophaga canimorsus endocarditis. In the case of culture negative endocarditis, it is important to carefully research animal contact and to inform the microbiology laboratory of the possibility of unusual bacteria such as Capnocytophaga canimorsus. There is no consensus about the choice of antibiotics and the length of treatment. Antibiotic therapies are often initiated empirically as Capnocytophaga canimorsus are slow growing bacteria. Most patients are successfully treated with broad-spectrum antibiotics as many strains are still susceptible to all antibiotics. High dose of β lactams remains a treatment of choice also there is a high rate of resistance to these. The efficacy of antibiotic therapy, must be confirmed after isolation and in vitro susceptibility testing of the strain. The difficulty in diagnosing Capnocytophaga canimorsus endocarditis may delay diagnosis and appropriate treatment and may explain the high mortality rate of 25% in spite of its low virulence. In conclusion, Capnocytophaga phage canimorsus produces rarely reported endocarditis whose incidence may be underestimated, considering its failure to grow on standard media. The combination of animal contact, underlying disease or predisposition to endocarditis and apparently culture negative endocarditis should raise the suspicion of Capnocytophaga canimorsus infection.

References