Riboflavin alleviates cardiac failure in Type I diabetic cardiomyopathy

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Abstract

Heart failure (HF) is a common and serious comorbidity of diabetes. Oxidative stress has been associated with the pathogenesis of chronic diabetic complications including cardiomyopathy. The ability of antioxidants to inhibit injury has raised the possibility of new therapeutic treatment for diabetic heart diseases. Riboflavin constitutes an essential nutrient for humans and animals and it is an important food additive. Riboflavin, a precursor of flavin mononucleotide (FMN) and flavin adenine dinucleotide (FAD), enhances the oxidative folding and subsequent secretion of proteins. The objective of this study was to investigate the cardioprotective effect of riboflavin in diabetic rats. Diabetes was induced in 30 rats by a single injection of streptozotocin (STZ) (70 mg/kg). Riboflavin (20 mg/kg) was orally administered to animals immediately after induction of diabetes and was continued for eight weeks. Rats were examined for diabetic cardiomyopathy by left ventricular (LV) remodynamic function.

Introduction

Cardiovascular disease is the most common serious complication of diabetes mellitus. Prominent defects of diabetic cardiomyopathy include the prolonged duration of contraction and relaxation1 and reduced cardiac compliance. Although diabetic cardiomyopathy has been the subject of intensive investigation over the last 30 years, the pathogenesis of diabetic cardiomyopathy is still far from being fully clarified. Various studies have reported that the circulating markers of myocardial damage, pro-inflammatory cytokines, calcium dysregulation, oxidative stress and atherogenic lipids are elevated in diabetic patients.2,9

Metabolically, the diabetic heart is characterized by diminished glucose utilization and increased fatty acid oxidation resulting in lipid accumulation in the myocardium.10,11 This myocardial lipotoxicity results in alterations in the inflammatory cytokine levels and evokes a cascade of disparaging changes that leads to cardiac damage.

Numerous studies have demonstrated that the pivotal mediator for the pathogenesis of diabetes and its cardiovascular complications is oxidative stress.12,13 The increase in the level of oxidative stress results from an increased generation of reactive oxygen species (ROS) or from a reduction in antioxidants, which was thought to contribute to the initiation and progression of cardiac dysfunction and remodeling of the extracellular matrix in the heart.14-17 ROS are continuously produced in most cells under physiological conditions, and their levels are regulated by a number of antioxidant enzymes, such as superoxide dismutase, glutathione peroxidase and catalase, as well as by other non-enzymatic antioxidants. The ability of antioxidants to inhibit these injuries has raised the possibility of new therapeutic treatment for diabetes and its complication. Heme oxygenase-1 (HO-1) is a ubiquitously expressed stress inducible enzyme that catalyzes heme into bilirubin, carbon monoxide (CO) and iron.18 The byproducts of heme catabolism exert pleiotropic cytoprotective effects in the heart. Bilirubin is a powerful antioxidant,19 and CO exerts vasodilatory, anti-inflammatory and anti-proliferative effects.20 On another level, antioxidant administration has been reported to show beneficial effects on parameters of oxidative stress and cardiovascular functions in experimental diabetes.21

Connective tissue growth factor (CTGF), cysteine-rich glycoproteins, is an important stimulant of fibrosis,22 and is currently suggested to be an important downstream amplifier of the effects of the profibrogenic master cytokine transforming growth factor (TGF)-β.23,24 For cardiac fibrotic responses to diabetes, attention has been paid recently to the role of CTGF, since several studies indicated that renal and cardiac fibrosis may not be fully dependent on TGF-β, but may be predominantly dependent on CTGF.25,26 Therefore, an ideal approach to prevent DCM may target reducing CTGF expression.

Riboflavin is a precursor of flavin mononucleotide (FMN) and flavin adenine dinucleotide (FAD), which serve as coenzymes for numerous oxidases and dehydrogenases in
Materials and Methods

Experimental animals

Sprague-Dawley rats weighing 200±20 g were provided by the Experimental Animal Center in Wannan Medical College and housed in a standard animal facility under controlled environmental conditions at room temperature 22±2°C and 12-h light-dark cycle. All experiments were performed in accordance with the Guide for the Care and Use of Laboratory Animals of the Chinese National Institutes of Health.

Induction of diabetes

Diabetes was induced by a single i.p. injection of STZ (70 mg/kg) dissolved in 0.1M citrate buffer (pH 4.5) in male Sprague-Dawley rats (180-220 g). The control group rats were treated with the same volume citrate buffer. Diabetes was confirmed at 72 h after STZ injection by measuring the glucose concentrations of peripheral blood obtained from the tail vein (One Touch SureStep Meter, LifeScan, CA, USA) and weekly thereafter. Insulin was not administered, and all animals had free access to food and water. Diabetes was diagnosed by a sustained glucose concentration of more than 15 mmol/l.

Experimental protocols and riboflavin treatment

Control and diabetic rats were randomly assigned to three main experimental groups (8 animals per group) immediately after confirmation of STZ-induced diabetes: control rats received the control diet; diabetic rats, received the control diet; riboflavin rats, received orally administered riboflavin (20 mg kg⁻¹ per day) and the control diet.

Cardiac function

After eight weeks of drug dosing, rats were anesthetized with sodium pentobarbital (50 mg/kg i.p.). The right carotid artery was cannulated with a Millar minature catheter and advanced into the aorta to record arterial pressure. The aortic catheter was then advanced into the left ventricle to record left ventricular systolic pressure (LVSP), left ventricular developed pressure (LVDP), left ventricular end diastolic pressure (LVEDP), maximal rate of rise/fall left ventricle pressure development and decline (±dP/dtmax). All pressure data were recorded on MedLab data acquisition system (Nanjing MedEase Co., Nanjing, China). Fasting blood samples and hearts were then collected from all the groups of rats for further studies.

Biochemical analyses of blood samples

The levels of serum triacylglycerol (lipoprotein lipase method), cholesterol (cholesterol oxidase method), HDL (polyanion polymer/detergent method), LDL (catalase method), creatine kinase (CK; peroxidase method) and lactate dehydrogenase (LDH; lactic acid method) were measured as previously described.

Measurement of myocardial oxidative stress

Left ventricular homogenate (10%, w/v) was prepared with 0.1 M PBS and centrifuged at 12,000 g for 10 min. The supernatant was used to determine SOD activity and MDA levels with commercially available kits (Nanjing Jiancheng Bioengineering Institute).

Western blotting

Left ventricles (0.2 g) were lysed and homogenized in 2 mL of lysis buffer (10 mM Tris-buffered saline, 1 mM EDTA, 1 mM EGTA, 2 mM sodium orthovanadate, 0.2 mM PMSF, 2 μg/mL leupeptin, 2 μg/mL aprotinin, and 1% Triton X-100) for 30 min on ice and cleared by centrifugation at 12,000 g for 15 min at 4°C. Total protein concentration was determined in the supernatant using the Bradford assay (Bio-Rad Laboratories, Hercules, CA, USA). For each lane, equal amounts of protein were mixed with sodium dodecyl sulfate (SDS) sample buffer and boiled for 5 min. Samples were separated on a 10% sodium dodecyl sulfate-polyacrylamide gel and then transferred to 0.2-μm nitrocellulose membrane. Nitrocellulose blots were blocked by incubation in TBST (10 mM Tris-HCl, pH 7.5, 150 mM NaCl, and 0.1% Tween 20) containing 5% non-fat milk for 1 h at room temperature and then incubated with a rabbit polyclonal anti-HO-1, CTGF, β-actin antibody (1:500 dilution; Wuhan Boster Biotechnologies, Wuhan, China) overnight at 4°C. After 3 washing steps, a secondary anti-rabbit antibody (1:10,000 dilution; Sigma Chemical Co, St. Louis, MO, USA) was added and incubated for 1 h. After being rinsed three times with wash buffer, the reaction was visualized by DAB (Bio Basic Inc., Canada). The relative amounts of the bands were quantified by densitometry using image software.

Results

Change in body weight and heart weight/body weight ratio after riboflavin treatment

After STZ administration, animals showed similar hyperglycemia in all experimental groups. There was no change in blood glucose levels in diabetic rats treated with riboflavin who also showed a significant decrease in body weight compared with controls (Table 1). However, the results in Table 1 also show an increase heart weight/body weight ratio in the STZ-diabetic animals. Treatment of riboflavin prevented body weight loss in diabetic rats as compared with the untreated rats. In addition, diabetic rats had increased heart weight/body weight ratio, a marker for the development of diabetic cardiomyopathy, and this ratio was significantly decreased by treatment with riboflavin (Table 1).

Effects of riboflavin on cardiac function

Cardiovascular function was evaluated by LV hemodynamic analysis before rats were sacrificed (Table 2). Left ventricular hemodynamic parameters were measured in control and dia-
Ripoflavin had a significant effect on lowering triglyceride, cholesterol and LDL levels and increasing HDL levels in diabetic rats (Table 3). All these levels were completely normalized after ripoflavin treatment which suggests that ripoflavin is far more effective in maintaining the lipid profile near to that of the control in this animal model of diabetes.

**Antioxidant effects of ripoflavin**

Lipid peroxide formation (MDA), which is a direct marker for oxidative stress, significantly increased the cardiac tissue of the STZ group compared with the control (Figure 2). Ripoflavin treatment significantly reduced the levels of lipid peroxidation which were very close to those of the control group (Figure 2). Treatment with ripoflavin significantly increased SOD heart activity in diabetic rats.

**Changes in HO-1, and CTGF protein by ripoflavin treatment in diabetic rats**

Western blot analyses showed an accumulation of HO-1 protein in diabetic rats compared with controls (Figure 2). The level of SOD was significantly depressed in the diabetic rats compared with controls (Figure 2). Treatment with ripoflavin significantly increased SOD heart activity in diabetic rats.
pared with normal control rats (Figure 3). Riboflavin treatment led to a strong upregulation of HO-1 protein compared with the untreated rats (Figure 3). To determine the effects of riboflavin on cardiac fibrotic response to diabetes, expression of cardiac CTGF was examined by Western blot. Results showed that CTGF was significantly increased in cardiac tissue from diabetic rats (Figure 3). Riboflavin treatment significantly prevented diabetes-caused upregulation of CTGF.

Discussion

Diabetes is a metabolic syndrome with a cluster of common clinical disorders that is related with an increased risk for cardiovascular disease.38 Cardiovascular complications remain the leading cause of diabetes-related mortality and morbidity.36 A specific disease termed as diabetic cardiomyopathy increases the risk for cardiac dysfunction and heart failure independently of other risk factors such as coronary artery disease and hypertension, as evidenced by compelling epidemiological and clinical data.37,38 Despite the potential importance of this disease entity, the underlying mechanisms are still not well understood. Dyslipidemia, oxidative stress and inflammatory injury are important interrelated factors responsible for the development of cardiomyopathy as they are known to promote the progression of premature atherosclerosis, coronary insufficiency and myocardial infarction.39 Riboflavin, a precursor of flavin mononucleotide (FMN) and flavin adenine dinucleotide (FAD), prevents cardiac damage and enhances the oxidative folding and subsequent secretion of proteins.40-42 However, the effect of riboflavin on the antioxidative properties and altered lipid profile in experimental models of diabetes has not yet been studied. In the present study, we extensively evaluated the effect of riboflavin on cardiac damage.

Diabetic cardiomyopathy characterized by diastolic dysfunction and left ventricular hypertrophy is usually the terminal condition of heart in diabetes.43 Our results indicate that 8-week treatment of riboflavin in diabetic rats proved to be beneficial as it restrained the progression of the metabolic disorders of diabetes. The STZ diabetic rats showed elevated blood glucose levels. Also, there was a notable increase in the HW/BW ratio which signifies cardiac hypertrophy. Metabolically, the diabetic heart is characterized by diminished glucose utilization and increased fatty acid oxidation resulting in lipid accumulation in the myocardium.10,11 Riboflavin remarkably improved lipid profile. Riboflavin treatment showed a distinct positive effect on HW/BW ratio indicating that riboflavin was able to prevent cardiac hypertrophy which usually sets in as a result of diastolic dysfunction secondary to diabetes. The untreated STZ-diabetic animals showed significant depressions in LVSP, +dP/dt, and −dP/dt and concomitant increases in LVEDP. These hemodynamic alterations in the untreated STZ-diabetic rats demonstrate abnormal left ventricular systolic and diastolic function that is the hallmark of diabetic cardiomyopathy. Riboflavin treatment attenuated these hemodynamic changes.

Increases in levels of circulating cardiac damage markers, such as CK and LDH, represent a powerful and sensitive predictor of increased cardiac complications.44-46 In the present study, riboflavin-treated diabetic rats showed a significant improvement in levels of these circulating cardiac damage markers. This indicates that riboflavin prevents cardiac damage and has beneficial properties. The decrease in the cardiac damage markers in the diabetic rats upon riboflavin treatment suggested that the riboflavin treatment reduces the risk of metabolic disorders associated with diabetes.

Increased oxidative stress and an altered antioxidant pool have been implicated in both clinical and experimental type 1 diabetes.14 The results from our study showed that an STZ induced diabetic condition resulted in an increase in the lipid peroxidation and protein carbonylation which are direct indicators of systemic oxidative stress. This was in conjunction with depletion of superoxide scavenger SOD and an increase in lipid peroxidation product MDA. Riboflavin treatment in the STZ-diabetic rats reduced the formation of lipid peroxides and carbonyl content and also restored the levels of SOD. Heme oxygenase-1 (HO-1), the inducible isoform of the HO system, is a rate-limiting enzyme which converts heme into equimolar amounts of iron, carbon monoxide and biliverdin. HO-1 is thought to have antioxidant and cytoprotective roles.31 Riboflavin treatment significantly increased the level of myocardial HO-1 in the STZ-diabetic rats. This positive beneficial effect of riboflavin could be directly attributable to its antioxidative nature which is in agreement with previous findings.40-42

Apart from the altered metabolic condition, diabetes is also an inflammation-prone condition. Hyperglycemia-induced ROS stimulates signal transduction to instigate inflammatory cytokines, e.g. TNF-α, IFN-γ and TGF-β.46 This leads to systemic inflammation, cardiac dysfunction and exacerbates the severity of diabetes.47 CTGF, a recently discovered cytokine, has been demonstrated to play an important role in fibrotic response through a TGF-β1-dependent or independent pathway.25,26 CTGF acts as a cofactor with TGF-β to induce fibroblasts to become myofibroblasts that deposit collagen, ultimately resulting in organ scarring and dysfunction, and in the most severe forms, organ failure and death. Indeed, CTGF levels in tissue, blood or vitreal fluid have been shown to correlate with the degree and severity of fibrosis in many diseases.48 Other studies showed that CTGF also exhibited prohypertrophic properties on cardiomyocytes.49 Furthermore, CTGF has been proposed as a heart failure biomarker.43-44 Stress induced CTGF in cultured cardiomyocytes.50,51 In the present study, riboflavin-treated diabetic rats showed effective suppression of CTGF. The results strongly indicate that riboflavin is a potential agent against diabetes-associated systemic fibrosis.

In our study, the altered lipid profile

![Figure 3. Western blot of HO-1 (a) and CTGF (b) in heart after treatment with riboflavin in diabetic rats. HO-1 and CTGF were normalized with respect to actin in respective controls. Similar results were obtained in four independent sets of experiments. All the values represent mean±SEM (n=4). **P<0.01, significant difference to control rats; ##P<0.01, significant difference to diabetic rats. HO-1, heme oxygenase-1; CTGF, connective tissue growth factor.](image-url)
observed in the diabetic condition was also abrogated by riboflavin treatment. Riboflavin improved the serum cholesterol, triglycerides, LDL and HDL. These findings suggest that riboflavin possesses lipid lowering activities due to its unique anti-atherogenic properties which contribute also to its cardiovascular protective actions. This is the first report to evaluate the positive effect of riboflavin on lipid profile in STZ-diabetic rats.

Limitations

This study has several limitations. The sample size was relatively small. This preliminary result must be verified in a more numerous population, and on a long-term basis. Although myocardial oxidative stress was measured, we did not provide more detailed mechanisms.

Conclusions

In summary, the present findings show that treatment with riboflavin may attenuate the progressive cardiac dysfunction and myocardial oxidative stress in a murine model of diabetic cardiomyopathy. The beneficial effect of riboflavin is a result of its inhibition of myocardial oxidative stress processes through its potential antioxidant properties. Our results show that riboflavin administration may exert beneficial effects in diabetic cardiomyopathy by increasing antioxidant and decreasing CTGF levels. We suggest that it might be useful to re-explore the role of antioxidant therapy given immediately after the diagnosis of type I diabetes mellitus to reduce the risk of future cardiovascular complications.

References

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[Heart International 2011; 6:e21]